

No.	K25-3165	
Project Title	Harnessing CRISPR-Cas9 and DNA Repair Pathways (COMBI-CRISPR) for Temporally Inducible IFNAR1 Mouse Models to Advance Vaccine Research for Severe Viral Diseases	
Principal Investigator	Silvia Marina Vidal (McGill University • Professor)	
Project Member(s)	IMSUT Host Researcher	Tomoji Mashimo (The Institute of Medical Science, The University of Tokyo • Professor)
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IMSUT International Joint Usage/Research Center Project <International>

Joint Research Report (Annual/Project Completion)

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Report
1. Communication and Collaborative Discussions We conducted a series of discussions with Dr. Silvia Vidal (McGill University; total of 8 exchanges), primarily focusing on the design strategy for mouse models (DC-SIGN and IFNAR1) and preparation for applications to the IMSUT Joint Research Program. Key milestones include: May 15, 2024: Scheduling coordination and Zoom meeting setup May 23, 2024: Receipt of publications and technical materials on hDC-SIGN mouse models November 26-30, 2024: Proposal and discussions on IFNAR suppression models related to dengue vaccine research, including a shift from a doxycycline-inducible system to the AID2 system In addition, we held technical consultations with Dr. Masato Kanemaki (National Institute of Genetics; total of 6 exchanges until September 2025), focusing on implementation of the AID2 system and access to mouse resources. These discussions provided detailed technical guidance, including: Recommendation of the AID2 system Availability of OsTIR1-expressing mouse lines from NIG/RIKEN BRC Mechanistic insights into degradation of membrane proteins such as IFNAR1 2. Design Strategy and Internal Coordination Based on the above discussions, a detailed technical design was shared with Prof. Mashimo on November 20, 2025, and the following strategy was agreed upon: System: AID2 (~250 bp; Addgene #140532) Knock-in design: Fusion of mAID2 tag to the C-terminal cytoplasmic region of IFNAR1 Resources: Introduction of OsTIR1-expressing mice from NIG or RIKEN BRC Induction method: Administration of synthetic auxin (5-Ph-IAA) via intraperitoneal injection or drinking water 3. Project Summary and Current Status This project aims to generate a novel IFNAR1 knock-in mouse model using the auxin-inducible degron (AID2) system, which enables rapid and precise protein degradation at the organismal level. Discussions initiated in May 2024 with Dr. Silvia Vidal on interferon receptor regulation led to a transition from a doxycycline-inducible system to the AID2-based post-translational control system, which offers superior responsiveness and reversibility. In September 2025, intensive consultations with Dr. Masato Kanemaki further refined the system through the adoption of mAID2 and 5-Ph-IAA, improving inducibility while minimizing background degradation in vivo. Based on these developments, the genome editing design was finalized to fuse the mAID2 tag to the C-terminal cytoplasmic region of IFNAR1. This system is expected to enable rapid and reversible depletion of IFNAR1 in specific tissues or developmental stages, thereby providing a powerful platform for dynamic functional analysis. At present, all preparatory steps prior to mouse generation have been completed, including gene sequence optimization, donor DNA design, and arrangements for obtaining OsTIR1-expressing mice. The next phase will involve electroporation into fertilized embryos using the COMBI-CRISPR technology developed in the Mashimo laboratory, with the goal of establishing the mouse line efficiently. The successful implementation of this approach is expected to provide a broadly applicable in vivo

platform for studying membrane proteins.

