

Center for Stem Cell Biology and Regenerative Medicine

Division of Stem Cell and Molecular Medicine

幹細胞分子医学分野

Professor	Atsushi Iwama, M.D., Ph.D.
Senior Assistant Professor	Motohiko Oshima, Ph.D.
Assistant Professor	Yaeko Nakajima, Ph.D.
Project Assistant Professor	Takako Yokomizo, Ph.D.
Project Assistant Professor	Shuhei Koide, Ph.D.
Project Assistant Professor	Ola Rizq, M.D., Ph.D.
Project Assistant Professor	Minori Tamai, M.D., Ph.D.

教授	博士(医学)	岩間厚志
講師	博士(医学)	大島基彦
助教	博士(医学)	中島やえ子
特任助教	博士(医学)	横溝貴子
特任助教	博士(医学)	小出周平
特任助教	博士(医学)	オラ リズク
特任助教	博士(医学)	玉井望雅

Stem cells have the remarkable capacity to both self-renew and give rise to many types of more specialized cells in the body, which explains their great therapeutic potential in regenerative medicine. But that's not the only reason stem cells have become such a hotbed of scientific inquiry. These cellular transformers also offer an invaluable research tool for probing the disease mechanisms that underpin cancer, aging and a host of other health problems. Our major interest is to elucidate the mechanisms of self-renewal and multi-lineage differentiation of hematopoietic stem cells (HSCs). We are also interested in how the deregulated HSC functions are associated with aging of our body and the development of age-related hematological malignancies. We approach these issues mainly from the view point of epigenetics.

1. Chromatin accessibility in stem cells unveils progressive transcriptional alterations in myelodysplastic syndrome

Motohiko Oshima^{1#}, Naoya Takayama^{2#}, Yaeko Nakajima-Takagi¹, Daisuke Shinoda¹, Naoki Itokawa¹, Shuhei Kurosawa¹, Satoshi Kaito¹, Takahiro Kamiya¹, Yuta Yamada¹, Shohei Andoh¹, Kensuke Kayamori^{1,3}, Sudip Kumar Paul², Maria Alejandra Kanashiro², Tomoya Muto³, Shokichi Tsukamoto³, Emiko Sakaida³, Eriko Sato⁴, Nozomi Yusa⁵, Kazuaki Yokoyama⁵, Yasuhito Nannya⁵, Seiya Imoto⁶, Bahiyar Rahmutulla⁷, Atsushi Kaneda^{7,8}, Kiyoshi Yamaguchi⁹, Yoichi Furukawa⁹, Noriko Doki¹⁰, Koji Eto², Kei Nishikawa¹¹, Ye Ding¹¹, Tomohiro Myojo¹¹, Yuka Harada¹², Hironori Harada¹³, Atsushi Iwama^{1,14}

¹Division of Stem Cell and Molecular Medicine, Center for Stem Cell Biology and Regenerative

Medicine, The Institute of Medical Science, The University of Tokyo, Tokyo, Japan. ²Department of Regenerative Medicine, Graduate School of Medicine, Chiba University, Chiba, Japan. ³ Department of Hematology, Chiba University Hospital, Chiba, Japan. ⁴ Department of Hematology, Juntendo University Nerima Hospital, Tokyo, Japan. ⁵ Department of Hematology and Oncology, Research Hospital, The Institute of Medical Science, the University of Tokyo. ⁶ Division of Health Medical Intelligence, Human Genome Center, The Institute of Medical Science, The University of Tokyo. ⁷ Department of Molecular Oncology, Graduate School of Medicine, Chiba University, Chiba, Japan. ⁸ Health and Disease Omics Center, Chiba University. ⁹ Division of Clinical Genome Research, Advanced Clinical Research Center, The Institute of Medical Science, The University of Tokyo. ¹⁰ Hematology Division, Tokyo Metropolitan Cancer and Infectious Diseases

Center, Komagome Hospital, Tokyo, Japan. ¹¹ Department of Oncology and Hematology, Edogawa Hospital, Tokyo, Japan. ¹² Department of Clinical Laboratory, Tokyo Metropolitan Cancer and Infectious Diseases Center, Komagome Hospital, Tokyo, Japan. ¹³ Laboratory of Oncology, School of Life Sciences, Tokyo University of Pharmacy and Life Sciences, Tokyo, Japan. ¹⁴ Laboratory of Cellular and Molecular Chemistry, Graduate School of Pharmaceutical Sciences, The University of Tokyo, Tokyo, Japan.

Myelodysplastic syndrome (MDS) originates from hematopoietic stem cell (HSC) clones with acquired gene mutations. However, the molecular characteristics of MDS stem cells remain poorly understood. Here, we show that the chromatin accessibility profiles of MDS stem cells more accurately reflect disease status than those of progenitor cells and reveal the process of stem cell alterations during disease progression. Characterization of differentially accessible regions (DARs) showed that MDS stem cells acquire progenitor-like chromatin accessibility during disease progression, leading to disruption of the normal stem-progenitor hierarchy. Profiling of transcriptional factor-binding motifs at DARs further uncovered precocious activation of myeloid transcriptional networks in MDS stem cells, with a concurrent loss of HSC-associated regulatory programs. In particular, increased chromatin accessibility at CEBP target sites represented the myeloid reprogramming status of MDS stem cells. Newly developed “progenitor scores” based on chromatin accessibility stratified disease status and correlated well with prognosis. These findings indicate that chromatin landscapes of MDS stem cells define their cell-autonomous behavior and contribute to disease progression.

2. Identification of a CD138-negative therapy-resistant subpopulation in multiple myeloma with vulnerability to splicing factor inhibition

Takahiro Kamiya^{1,2}, Masahiko Ajiro³, Motohiko Oshima¹, Shuhei Koide¹, Yaeko Nakajima-Takagi¹, Kazumasa Aoyama¹, Akiho Tsuchiya¹, Satoshi Kaito¹, Naoki Itokawa¹, Ryoji Ito⁴, Kiyoshi Yamaguchi⁵, Yoichi Furukawa⁵, Bahityar Rahmutulla⁶, Atsushi Kaneda^{6,7}, Takayuki Shimizu^{2,8}, Noriko Doki⁹, Taku Kikuchi¹⁰, Nobuhiro Tsukada¹⁰, Masayuki Yamashita^{1,11}, Shinichiro Okamoto², Akihito Yoshimi³, Keisuke Kataoka^{2,12}, and Atsushi Iwama^{1,13,14*}

¹ Division of Stem Cell and Molecular Medicine, Center for Stem Cell Biology and Regenerative Medicine, The Institute of Medical Science, The University of Tokyo, Tokyo, Japan. ² Division of Hematology, Department of Medicine, Keio University School of Medicine, Tokyo, Japan. ³ Division of Cancer RNA Research, National Cancer Center Research Institute, Tokyo, Japan. ⁴ Central Institute

for Experimental Medicine and Life Science, Kanagawa, Japan. ⁵ Division of Clinical Genome Research, Advanced Clinical Research Center, The Institute of Medical Science, The University of Tokyo, Tokyo, Japan. ⁶ Department of Molecular Oncology, Graduate School of Medicine, Chiba University, Chiba, Japan. ⁷ Health and Disease Omics Center, Chiba University, Chiba, Japan. ⁸ Division of Hematology, Department of Medicine, National Hospital Organization Tokyo Medical Center, Tokyo, Japan. ⁹ Hematology Division, Tokyo Metropolitan Cancer and Infectious Diseases Center, Komagome Hospital, Tokyo, Japan. ¹⁰ Department of Hematology, Japanese Red Cross Medical Center, Tokyo, Japan. ¹¹ Division of Experimental Hematology, Department of Hematology, St. Jude Children's Research Hospital, Memphis, TN, USA. ¹² Division of Molecular Oncology, National Cancer Center Research Institute, Tokyo, Japan. ¹³ Laboratory of Cellular and Molecular Chemistry, Graduate School of Pharmaceutical Sciences, The University of Tokyo, Tokyo, Japan.

The molecular basis of therapy resistance in multiple myeloma (MM) remains poorly understood. Here, we performed single-cell RNA sequencing coupled with VDJ-targeted sequencing of highly purified primary MM cells from patient bone marrow. This approach uncovered cellular heterogeneity and phenotypic plasticity of MM cells across a spectrum of CD138 expression, accompanied by drastic epigenetic alterations. Notably, therapy-resistant subpopulations were identified within a minor fraction of CD138⁺ MM cells, which were shown via CRISPR/Cas9 screening to be vulnerable to splicing pathway inhibition. Consistently, this fraction of CD138⁺ MM cells showed increased differential splicing associated with overexpression of SR protein family splicing factors. Among these splicing factors, RNA-binding protein 39 (RBM39) was overexpressed in therapy-resistant cells and involved in aberrant splicing. Both genetic and pharmacological RBM39 inhibition exhibited a significant lethal effect on MM cells. Collectively, our findings identify distinct therapy-resistant MM subpopulations and highlight targeting the splicing pathway as a promising therapeutic strategy.

3. Tracking *Clusterin* expression in hematopoietic stem cells reveals their heterogeneous composition across the lifespan

Shuhei Koide¹, Motohiko Oshima¹, Takahiro Kamiya¹, Zhiqian Zheng¹, Zhaoyi Liu¹, Ola Rizq¹, Akira Nishiyama², Koichi Murakami^{2,3}, Yuta Yamada¹, Yaeko Nakajima-Takagi¹, Bahityar Rahmutulla⁴, Atsushi Kaneda^{4,5}, Kazuaki Yokoyama⁶, Nozomi Yusa⁷, Seiya Imoto⁸, Fumihito Miura⁹, Takashi Ito⁹, Tomohiko Tamura^{2,3}, Claus Nerlov¹⁰, Masayuki Yamashita^{1,11}, Atsushi Iwama^{1,12,13}

¹ Division of Stem Cell and Molecular Medicine, Center for Stem Cell Biology and Regenerative Medicine, The Institute of Medical Science, The University of Tokyo, Tokyo, Japan. ² Department of Immunology, Yokohama City University Graduate School of Medicine, Yokohama, Japan. ³ Advanced Medical Research Center, Yokohama City University, Yokohama, Japan. ⁴ Department of Molecular Oncology, Graduate School of Medicine, Chiba University, Chiba, Japan. ⁵ Health and Disease Omics Center, Chiba University, Chiba, Japan. ⁶ Department of Hematology/Oncology, Research Hospital, The Institute of Medical Science, The University of Tokyo, Tokyo, Japan. ⁷ Department of Laboratory Medicine, Research Hospital, The Institute of Medical Science, The University of Tokyo, Tokyo, Japan. ⁸ Division of Health Medical Intelligence, Human Genome Center, The Institute of Medical Science, The University of Tokyo, Tokyo, Japan. ⁹ Department of Biochemistry, Kyushu University Graduate School of Medical Sciences, Fukuoka, Japan. ¹⁰ MRC Molecular Haematology Unit, MRC Weatherall Institute of Molecular Medicine, Radcliffe Department of Medicine, University of Oxford, Oxford, UK. ¹¹ Division of Experimental Hematology, Department of Hematology, St. Jude Children's Research Hospital, Memphis, TN 38105, USA. ¹² Laboratory of Cellular and Molecular Chemistry, Graduate School of Pharmaceutical Sciences, The University of Tokyo, Tokyo, Japan. ¹³ The University of Tokyo Pandemic Preparedness, Infection and Advanced Research Center (UTOPIA), The University of Tokyo, Tokyo, Japan.

Hematopoietic stem cells (HSCs) exhibit signifi-

cant age-related phenotypic and functional alterations. Although single-cell technologies have elucidated age-related compositional changes, prospective identification of aging-associated HSC subsets has remained challenging. In this study, utilizing *Clusterin* (*Clu*)-GFP reporter mice, we demonstrated that *Clu* expression faithfully marks age-associated myeloid/platelet-biased HSCs throughout life. *Clu*-GFP expression clearly segregates a novel age-associated HSC subset that overlaps with but is distinct from those previously identified using antibodies against aging marker proteins or reporter systems of aged HSC signature genes. *Clu*-positive (*Clu*⁺) HSCs emerge as a minor population in the fetus and progressively expand with age. *Clu*⁺ HSCs display not only an increased propensity for myeloid/platelet-biased differentiation but also a unique behaviour in the BM, favouring self-renewal over differentiation into downstream progenitors. In contrast, *Clu*-negative (*Clu*⁻) HSCs exhibit lineage-balanced differentiation, which predominates in the HSC pool during development but becomes underrepresented as aging progresses. Both subsets maintain long-term self-renewal capabilities even in aged mice but contribute differently to hematopoiesis. The predominant expansion of *Clu*⁺ HSCs largely drives the age-related changes observed in the HSC pool. Conversely, *Clu*⁻ HSCs preserve youthful functionality and molecular characteristics into old age. Consequently, progressive changes in the balance between *Clu*⁺ and *Clu*⁻ HSC subsets account for HSC aging. Our findings establish *Clu* as a novel marker for identifying aging-associated changes in HSCs and provide a new approach that enables lifelong tracking of the HSC aging process.

Publications

1. Yamagishi M, Iwama A, Kitabayashi I. Polycomb repressive complexes as therapeutic targets in hematologic malignancies. **Exp Hematol** 2025 Dec 22:105339. doi: 10.1016/j.exphem.2025.105339.
2. Oshima M, Takayama N, Nakajima-Takagi Y, Shinoda D, Itokawa N, Kurosawa S, Kaito S, Kamiya T, Yamada Y, Andoh S, Kayamori K, Paul SK, Kanashiro MA, Muto T, Tsukamoto S, Sakaida E, Sato E, Yusa N, Yokoyama K, Nannya Y, Imoto S, Rahmutulla B, Kaneda A, Yamaguchi K, Furukawa Y, Doki N, Eto K, Nishikawa K, Ding Y, Myojo T, Harada Y, Harada H, and Iwama A. Chromatin accessibility in stem cells unveils progressive transcriptional alterations in myelodysplastic syndrome. **Nat Commun** 16:10726, 2025.
3. Kamiya T, Ajiro M, Oshima M, Koide S, Nakajima-Takagi Y, Aoyama K, Tsuchiya A, Kaito S, Itokawa N, Ito R, Yamaguchi K, Furukawa Y, Rahmutulla B, Kaneda A, Shimizu T, Doki N, Kikuchi T, Tsukada N, Yamashita M, Okamoto S, Yoshimi A, Kataoka K, Iwama A. Identification of a CD138-negative therapy-resistant subpopulation in multiple myeloma with vulnerability to splicing factor inhibition. **Blood Cancer Discovery**, 6(6):602-622, 2025.
4. Koide S, Oshima M, Kamiya T, Zheng Z, Liu Z, Rizq O, Nishiyama A, Murakami K, Yamada Y, Nakajima-Takagi Y, Rahmutulla B, Kaneda A, Yokoyama K, Yusa N, Imoto S, Miura F, Ito T, Tamura T, Nerlov C, Yamashita M, and Iwama A. Tracking *Clusterin* expression in hematopoietic stem cells reveals their heterogeneous composition across the lifespan. **Blood** 146:62-75, 2025.
5. Ho AD, Iwama A. Special issue: Bone Marrow Aging. **Exp Hematol** 144:104750, 2025.
6. Manabe N, Sakurai T, Kihara-Negishi F, Nakazawa Y, Yamada T, Iwama A, Oikawa T. Antagonistic effects of PU.1 on Gfi-1B-induced erythroid colony formation in human cord blood cells. **Cell Mol Biol** 71(3):48-56, 2025.

7. Yogo T, Becker HJ, Kimura T, Iwano S, Kuchimaru T, Miyawaki A, Yokomizo T, Suda T, Iwama A, Yamazaki S. Progenitor Effect in the spleen drives early recovery via universal hematopoietic cell inflation. **Cell Rep** 44:115241, 2025.
8. Yamanashi Y, Takamaru S, Okabe A, Kaito S, Azumaya Y, Kamimura Y, Yamatsugu K, Kajirai T, Kurumizaka K, Iwama A, Kaneda A, Kawashima S, and Kanai M. Chemical catalyst manipulating cancer epigenome and transcription. **Nat Commun** 16:887, 2025.